

Review

Bambusa vulgaris: A comprehensive review of its traditional uses, phytochemicals and pharmacological activities

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1. Introduction

B. vulgaris is an erect, evergreen, clump-forming bamboo growing 15 - 20 metres tall (Figure 1). It grows in loose clumps that are free of thorns, has dark green leaves and lemon-yellow stems mostly with green stripes. The stems are initially tough, not straight or can split easily, stiff in nature, and have thick walls with narrow lanceolate leaves. The densely tufted stems are 4–10 cm thick and reach heights upto 10–20 m. The trunk can be flexible (alternately bent in various directions) or straight, drooping at the ends. The walls of the trunk are quite thick, and nodes grew marginally. The internodal segment is 20 to 45 cm. There may be sprouting of a few branches between the middle trunk nodes to the top (1). According to CABI (Invasive Species Compendium), new bamboo culms that emerge from the ground (bamboo sprouts) are the edible shoots of several bamboo species particularly *B. vulgaris* and *Phyllostachys edulis*. It's a vegetable that's used in a variety of Asian dishes and broths. They are sold in a variety of processed shapes and come in fresh, dried, and canned varieties (2).

Among all the different species of bamboo shoots available in the world, only some of them are edible. Out of the 136 species of bamboo that can be found in India, *B. pallida*, *B. tulda*, *B. polymorpha*, *B. balcooa*, *Dendrocalamus hamiltonii*, *Dendrocalamus giganteus*, and *Melocanna bambusoides* are the most popular edible bamboo species (3). The genera of bamboo shoots which are edible available in the USA are *Phyllostachys*, the important being *Phyllostachys dulcis*,
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Abstract

A versatile plant with many purposes, *Bambusa vulgaris* is primarily known for its industrial applications, but it is also becoming acknowledged as a possible source of bioactive substances and as a functional food. Every component of the bamboo plant, including the rhizome, culm shavings, leaves, roots, shoots, and seeds, has potential medical uses. This review aims to provide an insight into the traditional uses, and the various pharmacological activities exhibited by *B. vulgaris* extracts like analgesic, antihyperglycemic, antipyretic, anti-inflammatory, antioxidant, antimicrobial, antiviral, hepatoprotective, anti-amnesic, etc. It also has immense potential to be used as an important functional food as it has a high content of useful proteins, carbohydrates, high fiber content, and very low fat.

Keywords *Bambusa vulgaris*; Ethnopharmacology; Nutritive value; Bioactive compounds

Phyllostachys edulis, *Phyllostachys bambusoides*, *Phyllostachys pubescens*, *Phyllostachys nuda* and *Phyllostachys viridis* (4).

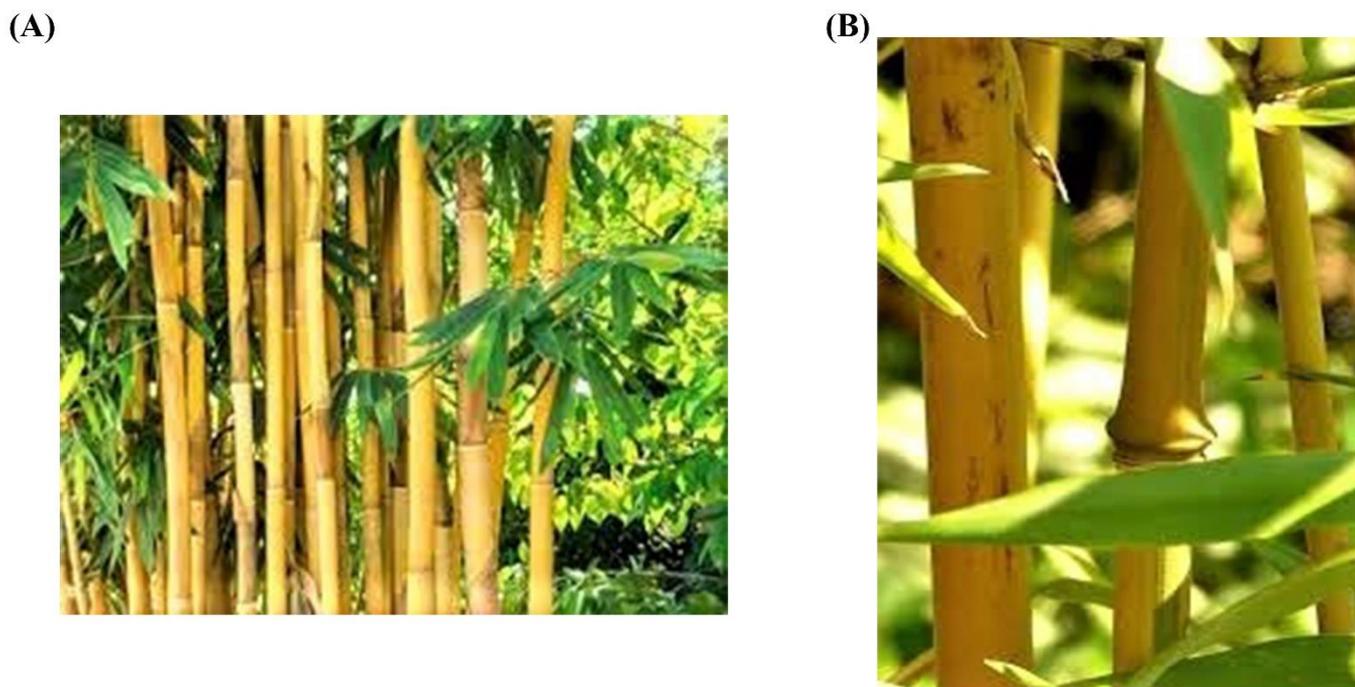


Figure 1 (A) Golden Bamboo (*B. vulgaris*) and (B) Close-up view of golden bamboo stems

2. Scientific classification

Kingdom: Plantae

Clade: Tracheophytes

Clade: Angiosperms

Clade: Monocots

Clade: Commelinids

Order: Poales

Family: Poaceae

Genus: Bambusa

Species: *B. vulgaris*

Binomial name: *Bambusa vulgaris*.

Synonym: *Bambusa auriculata*, *Gigantochloa auriculata*, *B. striata*

Local names: Baah Gaz (Assam), Tama (Nepal), Baseer Korool (Bengali).

3. Geographical location

Bamboo grows worldwide in at least 37 million hectares and covers 3.2 % of forest areas of their host countries or about 1 % of the global forest area. The southern tropical region of Asia contains about 80% of the area covered in bamboo. Bamboo is not very common in Africa or South America. In terms of the diversity of bamboo species, Madagascar has been named the richest nation in Africa. Eleven nations, one from Africa, eight from Asia, and two from Central and South America, contributed information to the Global Forest Resources Assessment 2000 (FRA 2000) on the size of their bamboo forests (5).

In Asia, Africa, and America bamboo is widely distributed. The Southeast Asia monsoon zone (south-eastern China, Indo-China, and the Indian subcontinent) is the world's bamboo distribution center; it is home to 90% of the world's total bamboo forest area as well as 80% of all bamboo species.

There are numerous bamboo species in both America and South Africa. It has to do with the history, architecture, and culture of some of these nations. The distribution centre for bamboo is located in Latin America, specifically in the



Amazon Basin near the South Tropic of Cancer, which includes Mexico, Guatemala, Costa Rica, Nicaragua, Honduras, Colombia, Venezuela, and Brazil. And the east coast of Madagascar is the centre of African bamboo distribution (6). India is the second richest country in the world, after China, in terms of bamboo genetic resources (5). The bamboo area of the country is estimated to be 15.69 million hectares with a total standing stock of 189 million tons (7).

4. Traditional Uses

Bamboo is inextricably linked to the cultural, social, and economic conditions of individuals in many Asian countries. It is the fastest-growing, multifunctional woody plant with a plethora of industrial and residential uses. Its use goes beyond just replacing wood in the building, furniture, scaffolding, and flooring; in China and South East Asia, it has traditionally been utilized as a source of food and medicine. The rhizome, culm, bark shavings, shoots, leaves, roots, and seeds of the bamboo plant are all used in medicine (42,43). Bamboo is currently attracting attention on a global scale for its nutritional and medicinal potential and is crucial to the food, pharmaceutical, and cosmeceutical industries. Bamboo shoots and leaves have excellent medicinal potential and can be used to treat illnesses naturally and sustainably (44, 45).

Bamboo has long been a crucial component of traditional Asian remedies, particularly Chinese and Indian (Ayurvedic) medications (45). Around 10,000 years ago, bamboo's medicinal uses were first documented in India for the preparation of Chyawanprash, a health tonic made from a variety of plants, including bamboo manna to promote youth, beauty, and longevity. Because of its ability to fight stress and slow the signs of aging, Chyawanprash has gained worldwide fame. The traditional Indian medical system of Ayurveda suggests using bamboo and its products, including Banslochan, Tabasheer, and Sitopaladi Churna, to cure a variety of illnesses (44,45). It is reported that in Pakistan, India, Brazil, and Tanzania it has been traditionally used as an astringent, emmanogogue, and abortifacient (47). Asase *et al.* (2010) reported that in Ghana it is used as a herbal remedy for the treatment of malaria (48). These modern pharmaceutical preparations made from bamboos, such as bamboo salt, bamboo starch, bamboo extracts, bamboo vinegar, bamboo silica, and more, are made using this ancient knowledge to address a variety of health issues like diabetes, inflammations, constipation, etc. (44,49,50).

5. Phytoconstituents present in *B. vulgaris*

Plants are the most abundant source of medications for ancient medical systems, modern medications, nutraceuticals, food supplements, folk remedies, pharmaceutical intermediates, and chemical entities for synthesised drugs (8). The ability to synthesise a wide range of chemical compounds allows plants to protect themselves from predators like insects, fungi, and herbivorous mammals as well as perform vital biological functions. Several active phytochemicals from plants were extracted and characterised to create a number of high-activity profile drugs (9). A variety of plant compounds and extracts have antioxidant or free radical-scavenging capabilities (10). These phytochemicals are separated into primary and secondary metabolites. Both the dry and wet ethanol-extracted leaf samples of *B. vulgaris* were subjected to a phytochemical analysis to determine their safety for ingestion (11). Table 1 presents the findings of the qualitative examination of *B. vulgaris*. All of the leaf extract was discovered to contain polyphenol and flavonoids in addition to saponin, general glycoside, coumarin, and cyanogenic glycoside. None of the species contained any remnants of anthraquinone, carotenoid, triterpenoid, steroid, or anthracene glycoside (12). In addition to the leaves and stems, some species of bamboo also have shoots that are valued for their health benefits due to their high protein, carbohydrate, vitamin, fibre, and mineral content and very low-fat content (12).

Table 1 Qualitative analysis of phytochemical constituents of *B. vulgaris*

PHYTOCHEMICAL	DETECTION	PLANT PART
Saponin	+	Leaf
Tannin	-	-
Terpenes	-	-
Flavonoid	+++	Leaf
Phlobatannins	-	-



Alkaloid	-	-
Glycosides	+	Ripe stem
Resin	+	Culm
Phenol	+	Leaf
Steroids	-	-
Proteins	+	Shoot
Carbohydrates	+	Shoot
Amino acids	+	Shoot
Gums & Mucilage	-	-
Non-reducing polysaccharides	-	-
Non-reducing simple sugar	+	Shoot

+ = mildly present; ++ = highly present; +++ = more highly present; - = absent or non-detectable.

5.1. Proteins

The protein content of bamboo shoots, which ranges between 1.49 g/100 g to 4.04 g/100 g and 21.1 g/100 g to 25.8 g/100 g on a wet and dry weight basis, is a potential source of proteins for humans. The species and maturity of the bamboo have a significant impact on the amount of protein in a bamboo shoot. The protein level of *B. vulgaris* was found to be 3.64 g/100 g (14). The composition of proteins in the bamboo shoot is of seventeen amino acids which are aspartic acid (Asp), serine (Ser), arginine (Arg), glutamic acid (Glu), threonine (Thr), glycine (Gly), proline (Pro), alanine (Ala), methionine (Met), valine (Val), histidine (His), cysteine (Cys), phenylalanine (Phe), lysine (Lys) leucine (Leu), isoleucine (Ile), and tyrosine (Tyr). All eight or at least seven essential amino acids (Lys, Ser, Met, His, Ile, Leu, Phe) for human beings are available in bamboo shoots (15, 51).

5.2. Carbohydrates

Bamboo shoots contained polysaccharides, oligosaccharides, and monosaccharides in terms of total carbohydrates. In bamboo shoots, the main polysaccharides are cellulose, hemicellulose, and starch, along with a few other minor complex polysaccharides like glycoproteins. Three oligosaccharides in particular—sucrose, arabinoxylan trisaccharide, tetrasaccharide, and xyloglucan disaccharide—were found to be the main ones in bamboo shoots. Dietary fibre with antioxidants is abundant in bamboo shoots. Bamboo shoots usually contained the monosaccharides fructose and glucose. The carbohydrate content of common species of newly emerged juvenile bamboo shoots usually ranges from 2.0 g/100 g to 9.94 g/100 g. (16).

5.3. Minerals

According to the results that are currently available, bamboo shoots are a good source of both macro and microelements. The main macro elements are potassium (K), phosphorus (P), sodium (Na), calcium (Ca), and magnesium (Mg), while the main microelements are cobalt (Co), copper (Cu), nickel (Ni), manganese (Mn), selenium (Se), iron (Fe), and zinc (Zn). Most studies found potassium to be the macroelement most abundant in bamboo shoots, followed by phosphorus and magnesium (17).



5.4. Vitamins

The majority of studies on vitamins have concentrated on vitamin C (ascorbic acid) and vitamin E (tocopherol). Vitamins C and E are intimately linked to the body's ability to produce antioxidants *in vivo*, but vitamin E synergistically with vitamin C strengthen the immune system. Fresh bamboo shoots contain far more vitamin C than vitamin E, which is also true of other common vegetables. Additionally, in some regions, fresh bamboo shoots are a respectable source of β -carotene and B-group vitamins. (18). The amounts of both vitamin C and vitamin E significantly dropped with the age of the shoots, according to a study by Nirmala *et al.* (2007). Additionally, the amount of vitamin C differed to a variable extent depending on the growth of bamboo shoots' altitude and distinct parts (tip and basal) (51).

5.5. Phenolic compounds

Bamboo shoots contained phenols that were primarily made up of flavonoids and phenolic acids. Bamboo shoots have been found to contain the following phenolic acids: protocatechuic acid, p-hydroxybenzoic acid, catechin, caffeic acid, chlorogenic acid, syringic acid, p-coumaric acid, ferulic acid, gallic acid, and vanillic acid (19). Protocatechuic acid, p-hydroxybenzoic acid, and syringic acid were the three most prevalent substances among them (20). There have been reports of fifteen phenolic acids, including 3-O-caffeoylshikimic acid, chlorogenic acid, p-coumaric acid, 3-p-coumaroylquinic acid, 5-p-coumaroylquinic acid, cryptochlorogenic acid, 1,3-dicaffeoyl quinic acid, 3,5-dicaffeoyl quinic acid, ferulic acid, 3-O-feruloylquinic acid, 5-O-ferul (52).

5.6. Flavonoids

Bamboo shoots and leaves contain flavonoids like orientin, isoorientin, isovitexin, vitexin, and tricetin (21). Bamboo tissues such as shoots, sheaths, and leaves mostly contained flavonoids in the insoluble form of free aglycone or flavonoid ligands. Apigenin 6,8-di-C-L-arabinopyranoside, 6-C-D-glucopyranosyl-8-C-L-arabinopyranosylchrysin, and kaempferide 3-O-L-rhamnopyranosyl were the seven flavonoids reported (1,6) 5,7,4'-trihydroxy-3',5'-dimethoxyflavone, narcissin, rutin, schaftoside, and D-glucopyranoside. The bamboo leaf has a flavone content of 2% to 5%, which has the ability to neutralise active free radicals, prevent sub-nitrification, and lower blood fat (52).

5.7. Phytosterols

Plants produce phytosterols in abundance, and *B. vulgaris* has been shown to be an excellent source of these compounds. To date, bamboo shoots have been found to contain seventeen phytosterols. Six phytosterols, including ergosterol, cholesterol, campesterol, stigmasterol, and β -sitosterol, were typically found in bamboo shoots. The total phytosterol content of bamboo shoots ranged from 66.60 mg/100 g to 242.77 mg/100 g on a dry basis, indicating the plant's ability to provide humans with useful phytosterols. Due to their wide range of health advantages, including their ability to decrease serum cholesterol and their anti-ulcer, anti-cancer, anti-inflammatory, and immunomodulatory properties, phytosterols were regarded as valuable dietary supplements (22).

5.8. Glycosides

The presence of cyanogenic glycosides has been reported in *B. vulgaris*. The cyanogen glycoside taxiphyllin is found in different levels in bamboo shoots (23-26). The β -glycosidase, which is produced in damaged bamboo shoot tissues, reacts with taxiphyllin to form dangerous hydrogen cyanide, whose concentration shouldn't be higher than what is toxic to humans (3). The majority of edible species of bamboo shoots have a significant quantity of cyanogen glycoside, with the shoot tip having the highest concentration. The detailed phytoconstituents that are found in *B. vulgaris* are provided in Table 2. The chemical structure of different phytoconstituents present in *B. vulgaris* is provided in Figure 2.

Table 2 Types of phytoconstituents present in *B. vulgaris*.

Classes	Phytoconstituents
Carbohydrates	Polysaccharides: cellulose, hemicellulose, Starch & glycoproteins. Oligosaccharides: sucrose, tetrasaccharide, arabinoxylan trisaccharide, xyloglucan disaccharide. Monosaccharides: fructose, glucose
Amino acids	Arginine (Arg), aspartic acid (Asp), serine (Ser), glycine (Gly), glutamic acid (Glu), alanine (Ala), threonine (Thr),



	proline (Pro), histidine (His), isoleucine (Ile), lysine (Lys) leucine (Leu), methionine (Met), cysteine (Cys), valine (Val), phenylalanine (Phe), and tyrosine (Tyr).
Peptides	Asp-Tyr (Dipeptide)
Minerals	Macroelements are mostly composed of Potassium (K), phosphorus (P), magnesium (Mg) calcium (Ca), sodium (Na), and phosphorus (P), whereas the majority of microelements mainly included cobalt (Co), copper (Cu), nickel (Ni), manganese (Mn), selenium (Se), iron (Fe) and zinc (Zn)
Vitamins	Vitamin B, Vitamin C, Vitamin E.
Phenols	Phenolic acids, p-hydroxybenzoic acid, protocatechuic acid, caffeic acid, catechin, chlorogenic acid, p-coumaric acid, syringic acid, gallic acid, ferulic acid, vanillic acid
Flavonoids	Apigenin 6,8-di-C- α -L-arabinopyranoside, kaempferide 3-O- α -L-rhamnopyranosyl (1,6)- β -D-glucopyranoside, 6-C- β -D-glucopyranosyl-8-C α -L-arabinopyranosylchrysin, narcissin, schaftoside, rutin, and 5,7,4'trihydroxy3',5'dimethoxyflavone.
Phytosterols	β -sitosterol, campesterol, stigmasterol, cholesterol, ergosterol and stigmastanol
Glycosides	Taxiphyllin (Cyanogenic glycoside)

6. Pharmacological Activity

6.1. Anti-inflammatory

A study was designed by Carey, W. M. *et al.* (2009) to investigate the anti-inflammatory effects of *B. vulgaris* methanol extract (MEBV) in mice. Acute inflammatory models such as formaldehyde-induced paw edema and acetic acid-induced vascular permeability were used to investigate anti-inflammatory effects, as were subacute anti-inflammatory models such as cotton pellet granuloma, plasma MDA estimation, and carrageenan-induced peritonitis (27).

In the formaldehyde-induced paw edema method, oral administration of MEBV in graded dosages (100, 200, and 400 mg/kg) resulted in a dose-dependent reduction in paw volume when compared to the control. The oral dose of 400 mg/kg had the considerable effects, resulting in a 46 % in paw volume ($P < 0.01$) when compared to the control. The anti-inflammatory activity at this dose was comparable to that of diclofenac (10 mg/kg, p.o.). The maximal anti-inflammatory impact was observed in all dosages of the test medication within 3 hours (27).

In the carrageenan-induced peritonitis model, MEBV decreased peritoneal leukocyte migration at rates of 38, 55.8, and 77.6 percent at doses of 100, 200, and 400 mg/kg, respectively, whereas indomethacin (10 mg/kg) inhibited it at a rate of 60.7 %. The neutrophil infiltration was inhibited by MEBV at 32.7, 54.3, and 64.9 %, respectively, whilst indomethacin inhibited it by 65.1 % (28).

6.2. Analgesic Activity

In acetic acid-induced writhing tests, the MEBV demonstrated dose-dependent and substantial analgesic efficacy. The administration of MEBV at dosages of 50, 100, 200, and 400 mg/kg reduced the number of writhing by 25.9 %, 29.6 %, 37.0 %, and 44.4 %, in experimental mice when compared to control group. These findings are comparable to those obtained when aspirin was administered to rats at doses of 200 and 400 mg/kg, which resulted in 40.7 % and 51.9 % reductions in writhing, respectively. Thus, at the highest dose of administration of MEBV 400 mg/kg, exhibited better analgesic activity than aspirin at 200 mg/kg (29).



6.3. Antipyretic activity

The antipyretic activities of *B. vulgaris* methanol extract has been investigated. The rectal temperature of experimental mice increased 18 hours after Brewer's yeast injection. In the study, a 1000 mg/kg dosage of *B. vulgaris* methanol extract shown considerable antipyretic efficacy. It produces a temperature drop after 2 hours, and by the end of the 5th hour, the temperatures of the two groups have gone back to normal (30).

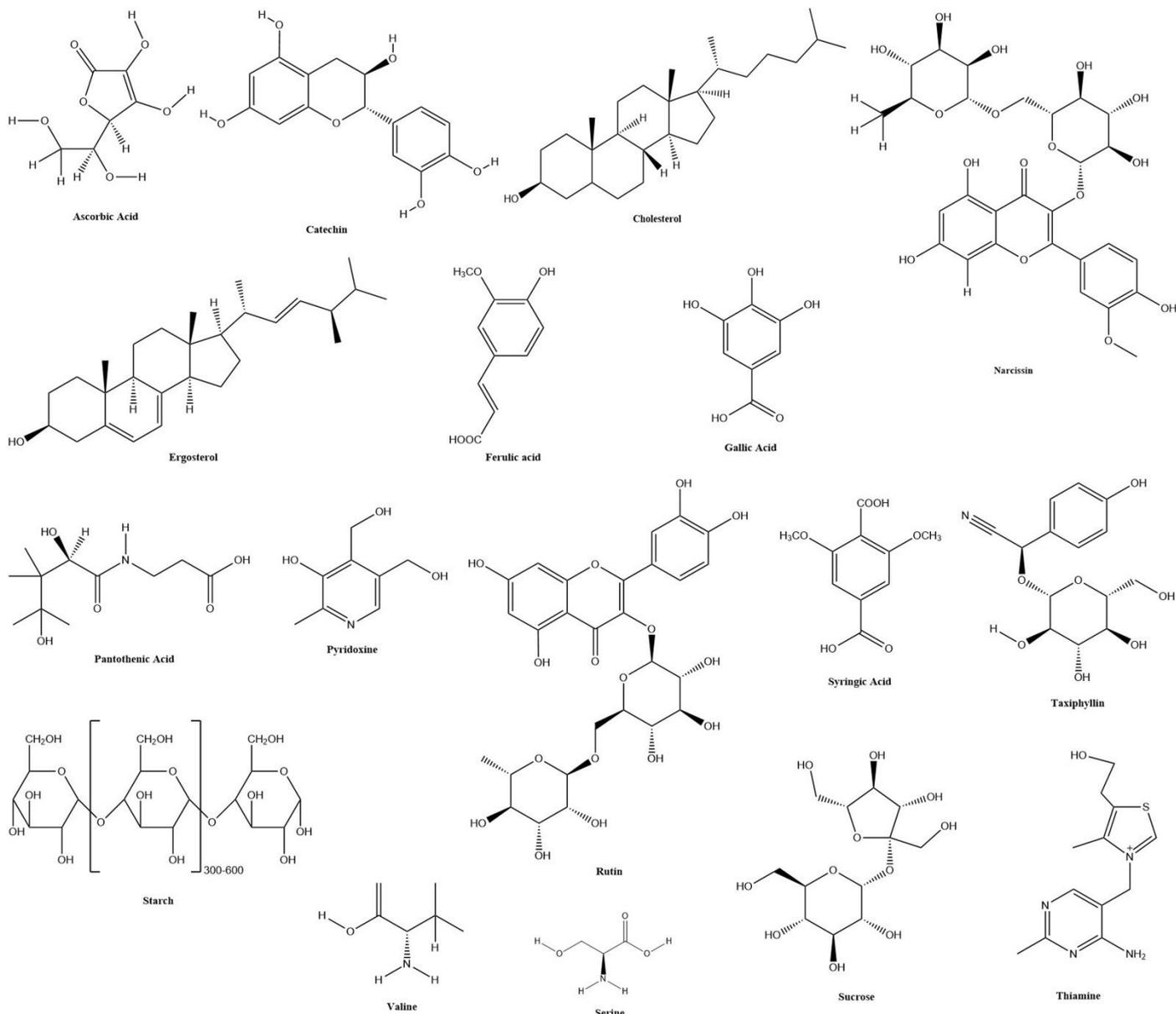


Figure 2 Structures of some phytoconstituents present in *B. vulgaris*.

6.4. Antihyperglycemic Activity

The methanol extract of *B. vulgaris* (MEBV) has exhibited good antihyperglycemic activity. MEBV, when given at doses of 50, 100, 200, and 400 mg/kg body weight in oral glucose tolerance tests, lowered the quantity of blood glucose in experimental animals' dose-dependently (30, 31). MEBV reduced blood glucose levels by 8.4, 32.8, 45.8, and 55.3 % at these four doses, respectively. However, at a dose of 50 mg MEBV per kg, the results were not statistically significant. Glibenclamide, a common antihyperglycemic drug, lowered blood glucose levels by 50.8 % when treated at a dose of 10 mg/kg body weight. Thus, at the highest dose studied, MEBV produced better antihyperglycemic activity to glibenclamide (30).



6.5. Antimicrobial Activity

The methanol, ethyl acetate and n-hexane extracts, of *B. vulgaris* can inhibit the growth of gram-positive and gram-negative bacteria and fungi. These extracts show strong antimicrobial activity against *Staphylococcus epidermidis* and *S. aureus*, *E. coli*, and *Aspergillus niger* (13,32). *B. vulgaris* methanol extract was evaluated against gram-positive, gram-negative, and fungi, *in vitro* investigation to assess its antimicrobial activity. Among gram-positive bacteria, maximum activity is exhibited against *B. subtilis*. On the other hand, the highest activity among gram-negative bacteria was seen in *E. coli*. Inhibition zones, particularly in kanamycin resistance, were reported to be 25 to 35 mm. When compared to the standard kanamycin, the zone of inhibition of the methanol extract was seen to be nearer to the standard (33). This study demonstrates the effectiveness of methanol extracts of *B. vulgaris* var. *Striata* against *S. aureus* and *E. coli*. It exhibits the largest zone of inhibition against *S. aureus* (with an average of 13.75 mm and 12.54 mm) and *E. coli* (with a mean of 8.64 mm and 8.86 12.54 mm) at 12-and 24-hours incubation. The presence of various phytochemicals in all *B. vulgaris* var. *Striata* extracts can be attributed for its antibacterial activity (34).

6.6. Hepatoprotective Activity

Anghore & Kulkarni (2016) investigated the hepatoprotective effect of the chloroform extract of leaves of *B. vulgaris*. The carbon tetrachloride-induced hepatotoxicity study in the liver cell of albino rats induces hepatic cell necrosis caused by metabolic activation and production of free radicals from CCl_4 . The administration of chloroform extract of *B. vulgaris* was found protective against CCl_4 -induced increase in enzyme levels of SGOT (Serum glutamic oxaloacetic transaminase), SGPT (Serum glutamic pyruvic transaminase), ALP (Alkaline phosphate) which served as a reliable pathological indicator for jaundice. There is a decrease in enzyme levels of SGOT, SGPT, and ALP which is comparable to the decreases in the standard group. Treatment with 200mg/kg body weight of chloroform extract of *B. vulgaris* reduced the elevation of SGOT, SGPT, and ALP. A dose of *B. vulgaris* extract of 250 mg/kg in albino rats was found to be potential against liver dysfunction and the dose was selected by LD_{50} for hepatoprotective activity (35).

6.7. Antioxidant Activity

Antioxidants can defend organisms from the oxidative stress caused by free radicals (1). According to a recent study, Satya *et al.*, (2009) investigated that fresh shoots of *B. vulgaris* have a 28.21% antioxidant activity when tested for their ability to scavenge free radicals by the DPPH method (36). The radical scavenging activity of chloroform, acetone, and methanol extract of the leaves of *B. vulgaris* from the DPPH assay, the IC_{50} values correspond to 389.23 $\mu\text{g/ml}$, 300.55 $\mu\text{g/ml}$, and 262.90 $\mu\text{g/ml}$ respectively (37).

6.8. Antimalarial activity

The decoction of *B. vulgaris* leaf is used traditionally by the people of Ghana for the treatment of malaria. Komlaga *et al.* (2016) studied the different extracts of *B. vulgaris* for their antiplasmodial activity against chloroquine-sensitive 3D7 *P. falciparum* and chloroquine-resistant W2 strain of *P. falciparum* where they found that the aqueous extract has shown moderate activity ($5 < \text{IC}_{50} < 15 \mu\text{g/ml}$) and the petroleum ether and ethyl acetate extracts have shown high activity ($\text{IC}_{50} < 1 \mu\text{g/ml}$) (38).

6.9. Anticonvulsant activity

The extract of leaves of *B. vulgaris* was investigated for anticonvulsant potential. Adebayo *et al.* studied the effect of the extract on the pentylenetetrazole-induced convulsion model and found at 100 mg/kg, 200 mg/kg, and 400 mg/kg exhibited ($p < 0.05$) prolongation of death time and offered 60%, 80%, and 100 % protection respectively compared to the control group (10ml/kg) which offered 0% protection. The dose of 400 mg/kg elongated the onset of clonic, tonic convulsions, and death latency (39).

6.10. Antiamnesic activity

The methanol extract of the leaves of *B. vulgaris* was studied for antiamnesic activity by scopolamine-induced amnesia on the Y-maze task. Scopolamine significantly ($p < 0.05$) reduced the percentage of correct alternation on the Y-maze when compared to the control-treated group on percentage alternation on the Y-maze task. However, when compared with the control-treated group, *B. vulgaris* extract significantly ($p < 0.05$) in a dose-dependent pattern increased the



reduced alternation induced by Scopolamine. Piracetam, a positive control drug significantly ($p < 0.05$) reversed the reduced alternation induced by Scopolamine in mice (39,40).

6.11. Antivirus

The ethanol extract of *B. vulgaris* was analyzed for its antiviral activity against three human viruses: measles, yellow fever, and poliovirus with standard laboratory tests of which the extract of *B. vulgaris* produces inhibition only for the measles virus at MIC 62.5 $\mu\text{g/mL}$ (41).

7. Conclusions

Bamboo has been used for centuries as a food source and to treat a variety of illnesses. It significantly affects people's socioeconomic well-being. Numerous research has evaluated the plant's potential as a medicine. Nevertheless, there is still a need for in-depth research on bamboo, outside of its application in food and crafts. Bamboo's ethnopharmacological uses must be backed up by substantial academic research before they can be widely used in a range of therapeutic procedures. Due to their high quantity of beneficial proteins, amino acids, carbs, and other essential minerals and vitamins, as well as their extremely low-fat content, they also have a significant potential for usage as crucial health foods.

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Conflict of Interest

The authors declare no conflicting interest.

Author contributions

All authors have equally contributed to the work.

References

1. Fitri, A., Asra, R., & Rivai, H. (2020). Overview of the traditional, phytochemical, and pharmacological uses of gold bamboo (*Bambusa Vulgaris*). *World Journal of Pharmacy and Pharmaceutical Sciences*, 9(8), 21.
2. Sangeetha, R., Diea, Y. K. T., Chaitra, C., Malvi, P. G., & Shinomol, G. K. (2015). The amazing bamboo: a review on its medicinal and pharmacological potential. *Indian J Nutr*, 2(1), 1-7.
3. Sharma, Y. M. L. (1980). Bamboos in the Asia Pacific Region. In *Bamboo research in Asia: proceedings of a workshop held in Singapore, 28-30 May 1980*. IDRC, Ottawa, ON, CA.
4. Rubatzky VE, Yamaguchi M (1997) *World vegetables: principles, production and nutritive values*. Chapman and Hall, New York, pp 658-660
5. Lobovikov, M., Paudel, S., Ball, L., Piazza, M., Guardia, M., Wu, J., & Ren, H. (2007). World bamboo resources: a thematic study prepared in the framework of the global forest resources assessment 2005 (No. 18). Food & Agriculture Org.
6. GEMCO energy (2022). Make bamboo pellets. <http://www.gemcopelletmills.com/make-bamboo-pellets.html>
7. Banerjee, S., Basak, M., Dutta, S., Chanda, C., Dey, S., Dey, A., Somkuwar, B. G., Kharlyngdoh, E., & Das, M. (2022). Sustainable uses of bamboo by indigenous people with special emphasis on North-East India. *Indigenous People and Nature*, 543-576. <https://doi.org/10.1016/B978-0-323-91603-5.00016-6>.
8. Hammer, K. A., Carson, C. F., & Riley, T. V. (1999). Antimicrobial activity of essential oils and other plant extracts. *Journal of applied microbiology*, 86(6), 985-990.
9. Mandal, V., Mohan, Y., & Hemalatha, S. (2007). Microwave assisted extraction—an innovative and promising extraction tool for medicinal plant research. *Pharmacognosy reviews*, 1(1), 7-18.
10. Larson, R. A. (1988). The antioxidants of higher plants. *Phytochemistry*, 27(4), 969-978.



11. Parekh, J., & Chanda, S. (2007). In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turkish Journal of Biology*, 31(1), 53-58.
12. Coffie, G. Y., Antwi-Boasiako, C., & Darkwa, N. A. (2014). Phytochemical constituents of the leaves of three bamboo (Poaceae) species in Ghana. *Journal of Pharmacognosy and phytochemistry*, 2(6).
13. Owolabi, M. S., & Lajide, L. (2015). Preliminary phytochemical screening and antimicrobial activity of crude extracts of *Bambusa vulgaris* Schrad. Ex JC Wendl.(Poaceae) from southwestern Nigeria. *American Journal of Essential Oils and Natural Products*, 3(1), 42-45.
14. Karanja, P. N. (2017). *Physicochemical properties of bamboo shoots of selected species grown in Kenya and utilization as human Food* (Doctoral dissertation, Faculty of Agriculture, JKUAT).
15. Lin, Z., Chen, J., Zhang, J., & Brooks, M. S. L. (2018). Potential for value-added utilization of bamboo shoot processing waste—recommendations for a biorefinery approach. *Food and bioprocess technology*, 11(5), 901-912.
16. Oshima, Y., Watanabe, T., Endo, S., Hata, S., Watanabe, T., Osada, K., & Takenaka, A. (2018). Effects of eicosapentaenoic acid and docosahexaenoic acid on anxiety-like behavior in socially isolated rats. *Bioscience, biotechnology, and biochemistry*, 82(4), 716-723.
17. Christian, A. L., Knott, K. K., Vance, C. K., Falcone, J. F., Bauer, L. L., Fahey Jr, G. C., ... & Kouba, A. J. (2015). Nutrient and mineral composition during shoot growth in seven species of *P hyllostachys* and *P pseudosasa* bamboo consumed by giant panda. *Journal of Animal Physiology and Animal Nutrition*, 99(6), 1172-1183.
18. Nirmala, C., Bisht, M. S., Bajwa, H. K., & Santosh, O. (2018). Bamboo: A rich source of natural antioxidants and its applications in the food and pharmaceutical industry. *Trends in Food Science & Technology*, 77, 91-99.
19. Pandey, A. K., & Ojha, V. (2013). Standardization of harvesting age of bamboo shoots with respect to nutritional and anti-nutritional components. *Journal of Forestry Research*, 24(1), 83-90.
20. Thapa, N., Lamichhane, J., & Gauchan, D. P. (2018). Phytochemical, antioxidant, antimicrobial and micropropagation study of *Bambusa* species. *Int. J. Res*, 5(21), 77-88.
21. Liu, H., Zhang, C., Liu, Y., & Duan, H. (2019). Total flavonoid contents in bamboo diets and reproductive hormones in captive pandas: exploring the potential effects on the female giant panda (*Ailuropoda melanoleuca*). *Conservation physiology*, 7(1), coy068.
22. Ogbe, R. J., Ochalefu, D. O., Mafulul, S. G., & Olaniru, O. B. (2015). A review on dietary phytosterols: Their occurrence, metabolism and health benefits. *Asian J. Plant Sci. Res*, 5(4), 10-21.
23. Nartey, F., Smith, R., & Bababumni, E. (1980). Toxicological aspects of cyanogenesis in tropical foodstuffs. *Toxicology in the Tropics*, 53-73.
24. Nongdam, P., & Tikendra, L. (2014). The nutritional facts of bamboo shoots and their usage as important traditional foods of northeast India. *International scholarly research notices*, 2014. Vetter J. Plant cyanogenic glycosides. *Toxicon*. 2000 Jan 1;38(1):11-36.
25. Schwarzmaier, U. (1977). Cyanogenesis of *Dendrocalamus*: taxiphyllin. *Phytochemistry*, 16(10), 1599-1600.
26. Seigler, D. S. (1991). *Herbivores: Their Interactions with Secondary Plant Metabolites*, edited by GA Rosenthal & MR Berenbaum.
27. Carey, W. M., Dasi, J. M. B., Rao, N. V., & Gottumukkala, K. M. (2009). Anti-inflammatory activity of methanolic extract of *Bambusa vulgaris* leaves. *International Journal of Green Pharmacy (IJGP)*, 3(3).
28. Griswold, D. E., Marshall, P. J., Webb, E. F., Godfrey, R., Newton Jr, J., DiMartino, M. J., ... & Hanna, N. (1987). SK&F 86002: a structurally novel anti-inflammatory agent that inhibits lipoxygenase-and cyclooxygenase-mediated metabolism of arachidonic acid. *Biochemical pharmacology*, 36(20), 3463-3470.
29. Haque, A. M., Das, A. K., Bashar, S. S., Al-Mahamud, R., & Rahmatullah, M. (2015). Analgesic and antihyperglycemic activity evaluation of *Bambusa vulgaris* aerial parts. *Journal of Applied Pharmaceutical Science*, 5(9), 127-130.
30. Senthil Kumar, M. K. (2012). *Pharmacognostical, Phytochemical and Pharmacological screening for Bambusa Vulgaris (Gramineae) and Pandanus Odoratissimus (Pandanaeae)* (Doctoral dissertation, CL Baid Metha College of Pharmacy, Chennai).
31. Joy, K. L., & Kuttan, R. (1999). Anti-diabetic activity of *Picrorrhiza kurroa* extract. *Journal of Ethnopharmacology*, 67(2), 143-148.
32. Naidu, M. A. (2012). Antimicrobial activity of methanolic extracts of bamboo shoots (*Bambusa vulgaris*). *International Journal of Pharmaceutical & Biological Archives*, 3(6), 1547-1549.
33. Rajeshwari, E. (2012). Evaluation of anti-microbial activity of *Bambusa vulgaris* leaves. *International Journal of Phytotherapy Research*, 2(2), 36-39.



34. Baguistan, B. J., Waing, K. G., & Valentino, M. J. (2017). Phytochemical Screening And Determination Of The Biological Activities Of *Bambusa vulgaris* Var. *Striata* And *Dendrocalamus asper* Shoot Extracts. *IJBPAS*, 6(11), 2109-2119
35. Anghore, D., & Kulkarni, G. T. (2016). Hepatoprotective effect of various extracts of *Bambusa vulgaris* *Striata* on Carbon tetrachloride-induced liver injuries. *International Journal of Pharmaceutical Research and Allied Sciences*, 5(3), 16-22
36. Satya, S., Singhal, S., Prabhu, G., Bal, L. M., and Sudhakar, P. (2009). Exploring the nutraceutical potential and food safety aspect of bamboo shoot of some Indian species. Proceedings of the World Bamboo Congress, 16–19 September, Bangkok
37. Singhal, P., Bal, L. M., Satya, S., Sudhakar, P., & Naik, S. N. (2013). Bamboo shoots: a novel source of nutrition and medicine. *Critical reviews in food science and nutrition*, 53(5), 517-534.
38. Komlaga, G., Cojean, S., Dickson, R. A., Beniddir, M. A., Suyyagh-Albouz, S., Mensah, M. L., ... & Loiseau, P. M. (2016). Antiplasmodial activity of selected medicinal plants used to treat malaria in Ghana. *Parasitology Research*, 115(8), 3185-3195.
39. Adebayo, M. A., Akinpelu, L. A., Okwuofu, E. O., Ibia, D. E., Lawson-Jack, A. F., & Igbe, I. (2020). Anticonvulsant, antiamnesic and anxiolytic activities of methanol leaf extract of *Bambusa vulgaris* (Poaceae) in mice. *Journal of African Association of Physiological Sciences*, 8(2), 149-157.
40. Akinpelu, L. A., Akanmu, M. A., & Obuotor, E. M. (2018). Mechanism of anticonvulsant effects of ethanol leaf extract and fractions of *Milicia excelsa* (Moraceae) in mice. *Journal of Pharmaceutical Research International*, 23(4), 1-11.
41. Ojo, O. O., Oluyeye, J. O., & Famurewa, O. (2009). Antiviral properties of two Nigerian plants. *Afr. J. Plant Sci*, 3(7), 157-159.
42. de Zoysa, N., Hettige, U., & Vivekanandan, K. (1988). Some aspects of bamboo and its utilization in Sri Lanka. *BAMBOOS*, 6.
43. Nirmala, C., Bisht, M. S., Bajwa, H. K., & Santosh, O. (2018). Bamboo: A rich source of natural antioxidants and its applications in the food and pharmaceutical industry. *Trends in Food Science & Technology*, 77, 91-99.
44. Nirmala, C., & Bisht, M. S. (2017). 10 WBC Reports: Bamboo: A prospective ingredient for functional food and nutraceuticals. *Bamboo journal*, (30), 82-99.
45. Tewari, D. N. (1988). Bamboo as poverty alleviator. *Indian Forester*, 114(10), 610-612.
46. Chongtham, N., Bisht, M. S., & Haorongbam, S. (2011). Nutritional properties of bamboo shoots: potential and prospects for utilization as a health food. *Comprehensive Reviews in Food Science and Food Safety*, 10(3), 153-168.
47. Bernstein, N., Akram, M., Yaniv-Bachrach, Z., & Daniyal, M. (2021). Is it safe to consume traditional medicinal plants during pregnancy?. *Phytotherapy Research*, 35(4), 1908-1924.
48. Asase, A., Akwetey, G. A., & Achel, D. G. (2010). Ethnopharmacological use of herbal remedies for the treatment of malaria in the Dangme West District of Ghana. *Journal of ethnopharmacology*, 129(3), 367-376.
49. Silva, M. F., Menis-Henrique, M. E., Felisberto, M. H., Goldbeck, R., & Clerici, M. T. (2020). Bamboo as an eco-friendly material for food and biotechnology industries. *Current Opinion in Food Science*, 33, 124-130.
50. Park, E. J., & Jhon, D. Y. (2009). Effects of bamboo shoot consumption on lipid profiles and bowel function in healthy young women. *Nutrition*, 25(7-8), 723-728.
51. Nirmala, C., David, E., & Sharma, M. L. (2007). Changes in nutrient components during ageing of emerging juvenile bamboo shoots. *International Journal of Food Sciences and Nutrition*, 58(8), 612-618.
52. Wang, Y., Chen, J., Wang, D., Ye, F., He, Y., Hu, Z., & Zhao, G. (2020). A systematic review on the composition, storage, processing of bamboo shoots: Focusing the nutritional and functional benefits. *Journal of Functional Foods*, 71, 104015.



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