


Research Article

Design and screening of tetracycline antibiotics: an *in-silico* approach

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Abstract: A prominent class of broad-spectrum antibiotics known as tetracycline works by inhibiting the synthesis of proteins, which prevents the development of bacteria. Tetracycline resistance is typically attributed to one or more of the following causes: ribosomal binding site mutations, acquisition of mobile genetic elements carrying tetracycline-specific resistance genes, and/or chromosomal mutations that increase the expression of intrinsic resistance mechanisms. In this research, our objective is to virtually plan and conduct in-silico experiments to find tetracycline derivatives with inhibitory capability against tetracycline resistance protein. The tetracycline derivatives were screened using the Data Warrior, Discovery Studio, PyRx, and Swiss ADME web tools. Initially, 19 tetracycline derivatives were primarily screened for ADME and toxicity study followed by docking study. Among the tetracycline derivatives, C1, C11, C12, C14, C16, and C17 were found to be the potential drug-like molecules with binding energies of -8.9 kcal/mol, -8.4 kcal/mol, -8.5 kcal/mol, -7.7 kcal/mol, -7.7 kcal/mol, -8.6 kcal/mol respectively. In particular, C1 was predicted to have a better binding affinity towards the target protein than the others.

1. Introduction

Tetracyclines are a significant group of broad-spectrum antibiotics that act by obstructing protein production to stop bacterial growth. Compounds of this broad family have bacteriostatic action and are used to treat a variety of bacterial infections, including Gram-positive and Gram-negative infections as well as diseases brought on by protozoan parasites and intracellular organisms (1). Tetracyclines have a hydronaphthacene nucleus and four linearly condensed benzene rings as their primary structural component.

Despite the notable success of penicillin, several industries, and academic institutions have been concerned with finding novel antibiotics synthesized by microorganisms by analyzing a large number of soil samples submitted from all over the world. Actinomycete bacteria were found to produce a yellow colony that had a remarkable inhibitory effect against a variety of pathogenic strains, including rickettsia and Gram-positive and Gram-negative bacteria. Aureomycin (syn. chlortetracycline) was the first tetracycline extracted from *Streptomyces aureofaciens*. With the isolation of *Streptomyces rimosus* by Pfizer, the antibiotics

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terramycin (syn. oxytetracycline) and aureomycin were obtained. Based on the chemical structure of Chlortetracycline by catalytic hydrogenation, tetracycline was discovered in 1953, with an improved pharmacokinetic profile. After discovering the first generation of tetracyclines (Chlortetracycline, oxytetracycline, and tetracycline), scientists have begun the development of newer tetracyclines with better pharmacokinetic properties, broader antimicrobial spectrum, and lower toxicity (2). The basic structure of tetracycline is given in figure 1, substitution on the different positions can lead to different compounds. The classification of a different generations of tetracyclines has shown in table 1.

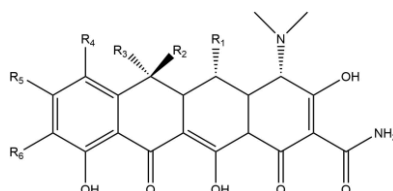


Figure 1 General structure of tetracycline

Table 1 Classification of the different generations of tetracyclines

Generations	Obtaining method	Drugs
First	Biosynthesis	Chlortetracycline, Oxytetracycline, Tetracycline, Demeclocycline
Second	Semi-synthesis	Doxycycline, Minocycline, Lymecycline, Meclocycline, Methacycline, Rolitetracycline
Third	Semi-synthesis Synthetic	Tigecycline, Omadacycline, Sarecycline Eravacycline

For extended spectrum β -lactamase (ESBL)-producing *Escherichia coli* and *Klebsiella* species (spp.), the prevalence of tetracycline resistance was found to be 66.9% and 44.9%, respectively. For methicillin-resistant *Staphylococcus aureus* (MRSA) and *Streptococcus pneumonia*, the prevalence was 8.7% and 24.3%, respectively (3). Tetracycline resistance is typically attributed to one or more of the following factors: chromosomal mutations that increase the expression of intrinsic resistance mechanisms, the acquisition of mobile genetic elements carrying tetracycline-specific resistance genes, and/or ribosomal binding site mutations. Computational approaches in drug design, discovery, and development process gain very rapid exploration, implementation, and administration. Introducing a new drug into a market is a very complex, risky, and costly process in terms of money and manpower. For reducing time, cost, and risk-born factors, the computer aided drug design (CADD) method is widely used as a new drug design approach (4). There are mainly two types of approaches for drug design through CADDs: Structure-based drug design/direct approach and Ligand-based drug design/ indirect approach. Docking is a valuable approach for studying the many features connected to protein-ligand interactions. In the field of molecular modelling, it predicts the preferred orientation of one molecule to another when they are linked together to form a stable complex. In this study, different compounds have been designed based on the general structure of the tetracycline and performed basic in-silico studies to find the best compound derivatives that can be analysed in resistant bacterial cells (5).

2. Materials and methods

2.1 Retrieval of target protein

The target site is the regulator (Tet R) of a membrane-associated protein (Tet A) that exports the antibiotic out of the bacterial cell before it attaches to the ribosomes and inhibits polypeptide elongation. The X-Ray crystal structure of Tet R was downloaded from RCSB-PDB in PDB file format with a PDB ID of 2TCT. The crystal structure of the target protein is shown in figure 2. The co-crystal inhibitor (CTC) was also identified and downloaded in SDF format.

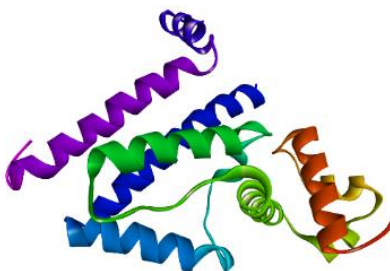


Figure 2 Crystal structure of Tet Repressor

2.2 Preparation of protein

The protein was prepared by the use of open-source BIOVIA Discovery Studio 2021 software (6). The water molecules and heteroatoms were also removed from the target protein. The active binding sites are defined with the 'Define and edit binding site' feature of the Discovery Studio software and the active site co-ordinates (x= 21.221424; y= 36.743182; z= 35.153182) were noted and saved for future use. Finally, polar hydrogen was added to the target protein and saved in PDB file format for future use.

2.3 Preparation of compound library

The structure of twenty compounds of tetracycline was prepared manually using Chem Draw Professional 16.0.0.82(68) software. The SMILES ID of these structures was also saved for future use. The compounds prepared with the software were saved in MDL SD File (*.SDF) format.

2.4 Molecular docking simulation studies

Molecular docking simulation studies using Autodock Vina on PyRx 0.8 virtual screening platform were conducted to predict the binding affinity between the target proteins and the produced drugs (7). The prepared protein was inserted into the 3D scene in the software's virtual platform and converted into PDBQT file format when processed into macromolecules (8). The downloaded proteins were reloaded. On enlarging Chain A of the corresponding protein, the amino acid sequence and the structure of the co-crystal ligand were made visible. To pinpoint precisely where the co-crystal inhibitors were located at the protein's binding site, the atoms of the co-crystal ligand were tagged. The pre-determined active binding site coordinates were utilized to modify the alignment of the 3D affinity grid box in the Vina search space of the PyRx 0.8 tool such that all of the amino acids are covered at the protein's active binding site. The default 3D affinity grid box size of 25 was maintained. And finally, MDSS was performed by the PyRx tool's predefined protocols (9).

2.5 Visualization and analysis of ligand interactions

Using the Discovery Studio Visualizer Software, the 2D interaction of the compounds with the highest binding affinities to the protein was viewed. Additionally, the co-crystal-protein ligand interactions in two

dimensions were seen. The compound was excluded from the investigation if it failed to create any regular hydrogen bonds with the active site residues.

2.6 ADMET Study

The ADME and toxicity prediction was done with Swiss ADME (10) and Data Warrior tools respectively. The SMILES ID of the compounds were entered in the required box and ADME and toxicity profiles (mutagenicity, carcinogenicity, reproductive toxicity, irritant) were generated.

3. Results

3.1 Features of the target protein and tetracycline derivatives

The RCSB-PDB website was used to get the crystal structure of protein chain A, shown in figure 3. The protein is made up of Chain A, which is complexed with co-crystal inhibitor viz. CTC (Chlortetracycline). The 2D chemical structure of nineteen tetracycline derivatives is given in figure 4.

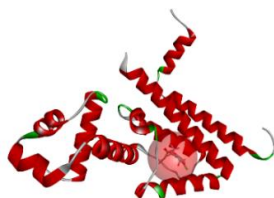


Figure 3 Chain A of 2TCT (Tet Repressor) with co-crystal inhibitor

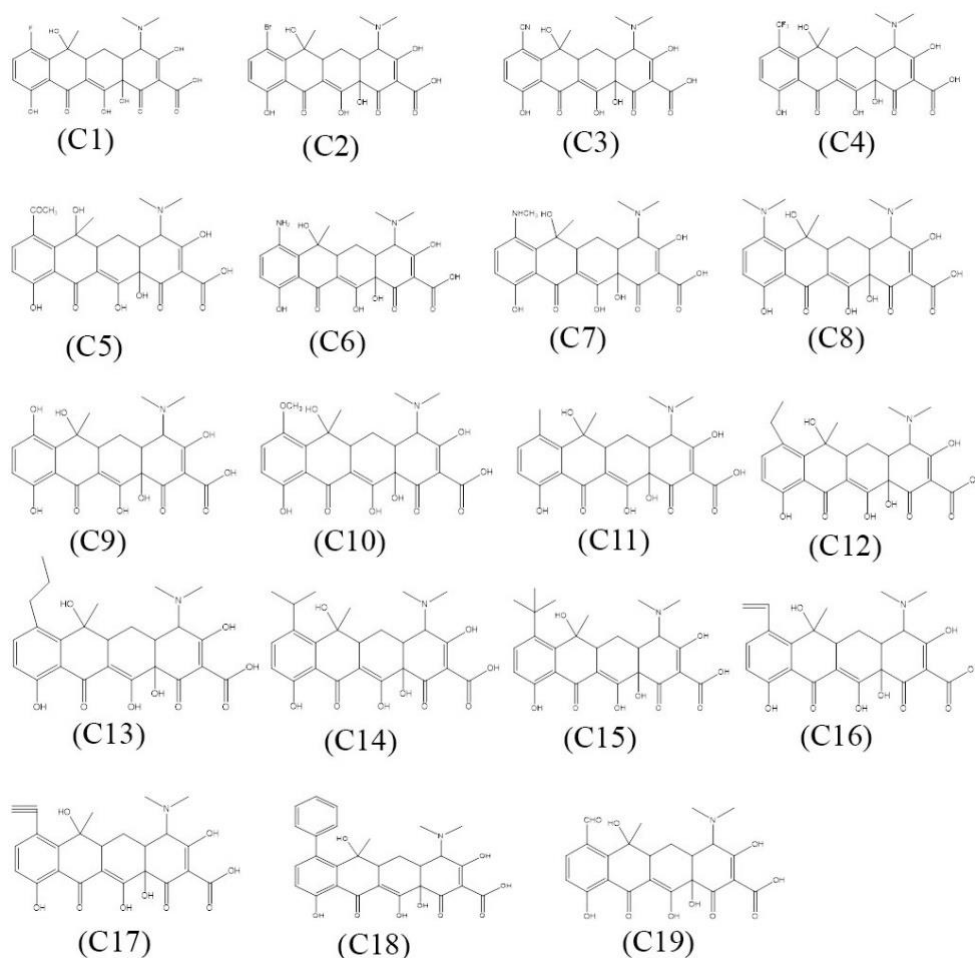


Figure 4 2D chemical structure of Tetracycline derivatives

3.2 ADMET analysis

The ADME characteristics and toxicity of the compounds were established after the binding affinity of the tetracycline derivatives was established. Due to weak ADME qualities, substances that are active in in-vitro environments may not be as active in-vivo. Therefore, it is necessary to research a compound's bioavailability. The Lipinski Rule of Five, also referred to as Pfizer's Rule of Five or simply the Rule of Five is a general guideline used to assess a chemical compound's drug-likeness or to determine whether it possesses chemical and physical characteristics that would likely make it an orally active drug in humans.

Drugs are frequently taken off the market from clinical use due to toxicity. Many researchers rely on Data Warrior, a trustworthy program, to forecast the toxicity of substances. We conducted an in-silico toxicity study, to ascertain the toxicity of the compounds. The ADME and toxicity study analysis is given in table 2.

Table 2 ADMET analysis of the compounds

Compound	Molecular weight (g/mol)	H-bond acceptor	H-bond donor	iLogP	Lipinski violation	Toxicity			
						Mutagenicity	Tumorigenicity	Reproductive	Irritant
C1	463.41	11	6	1.62	Yes	No	No	No	No
C2	524.32	10	6	2.04	No	No	No	No	No
C3	470.43	11	6	0.68	No	No	No	No	No
C4	513.42	13	6	1.48	No	No	No	No	No
C5	487.46	11	6	1.20	No	No	No	No	No
C6	460.43	10	7	0.25	No	No	No	No	No
C7	474.46	10	7	1.37	No	No	No	No	No
C8	488.49	10	6	1.93	No	No	No	No	No
C9	461.42	11	7	1.59	No	No	No	No	No
C10	488.49	10	6	1.93	No	No	No	No	No
C11	459.45	10	6	2.03	Yes	No	No	No	No
C12	473.47	10	6	2.02	Yes	No	No	No	No
C13	459.45	10	6	2.03	Yes	No	No	No	No
C14	487.50	10	6	2.33	Yes	No	No	No	No
C15	501.53	10	6	1.42	No	No	No	No	No
C16	471.46	10	6	2.05	Yes	No	No	No	No
C17	469.44	10	6	1.85	Yes	No	No	No	No
C18	521.52	10	6	2.35	No	No	No	No	No
C19	469.44	10	6	1.85	Yes	No	No	No	No

3.3 Molecular docking simulation studies

The molecular docking method enables us to define the behaviour of small molecules in the binding site of target proteins and to better understand basic biological processes by simulating the interaction between a small molecule and a protein at the atomic level. To evaluate the binding potential of ligands toward a protein, a docking study using the PyRx program can produce a binding affinity value (kcal/mol) for each ligand. Table 3 consists of a list of binding affinities of the compounds to the protein binding site.

Table 3 Binding affinities of the compounds with the protein binding site

Sl. No.	Compounds	Binding affinities (-kcal/mol)
1.	CTC (Co-crystal inhibitor)	7.7
2.	C1	8.9
12.	C11	8.4
13.	C12	8.5
14.	C13	7.5
15.	C14	7.7
17.	C16	7.7
18.	C17	8.6
20.	C19	7.6

3.4 Visualization and analysis of ligand interactions

Visualization of the 2D ligand interactions was done with Discovery Studio Visualizer software. The images of the 2D ligand interactions of the compounds with the Ter A protein are given in Figure 5. The summary of the ligand interactions of each drug with the amino acids of the protein is also given in Table 4.

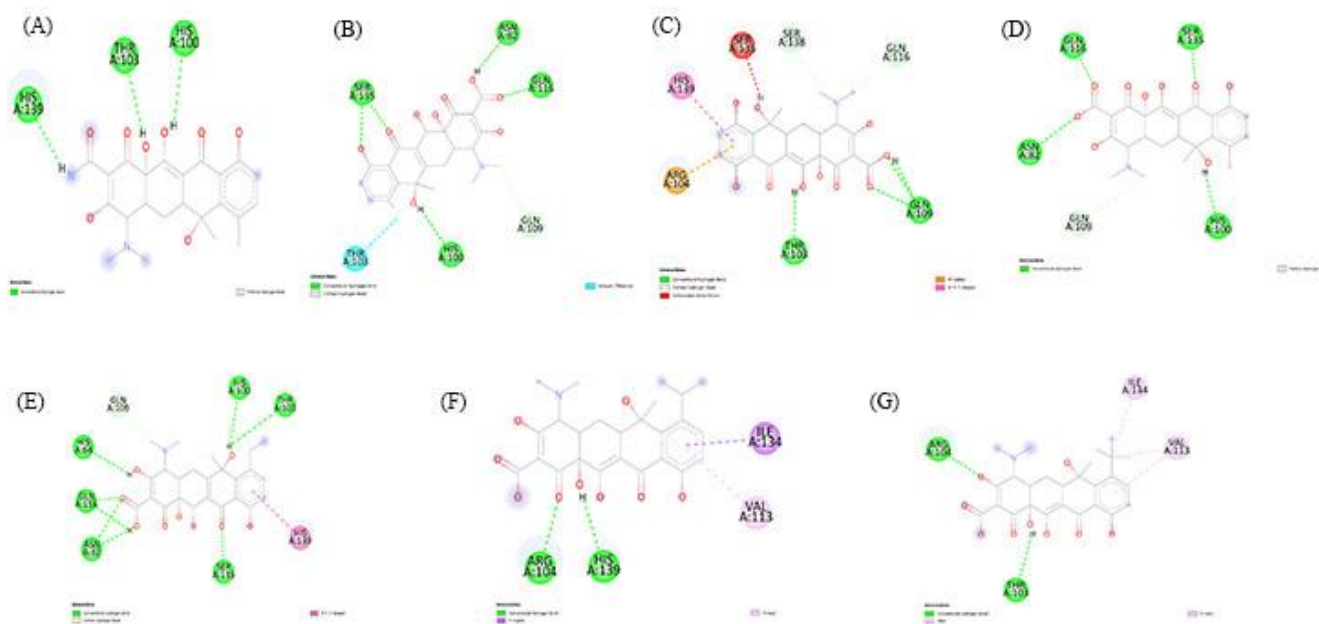


Figure 5 Visualization of 2D Interactions of the compounds with the Ter A protein: (A) CTC (Co-crystal inhibitor), (B) C1, (C) C11, (D) C12, (E) C14, (F) C16, (G) C17

Table 4 Summary of target-ligand interactions

Compounds	Conventional Hydrogen bond	Other binding sites
CTC (Co-crystal inhibitor)	HIS100 (2.70Å), HIS103 (2.15Å), HIS139 (2.23Å)	--
C1	ASN82 (2.09Å), GLN116 (1.71Å), HIS100 (2.40Å), SER135 (1.78Å, 2.54Å)	THR103 (2.79Å), GLN109 (3.58Å)
C11	ASN82 (2.57Å), HIS100 (2.29Å), GLN116 (1.75Å), SER135 (1.73Å)	GLN109 (3.51Å)

C12	HIS64 (2.91Å), ASN82 (2.62Å, 2.72Å), HIS100 (2.92Å), THR103 (3.00Å), GLN116 (1.78Å, 2.49Å), SER135 (1.80Å)	GLN109 (3.65Å), HIS139 (4.86Å)
C14	ARG104 (4.12Å), HIS139 (2.03Å)	VAL113 (4.49Å), ILE134 (3.90Å)
C16	GLY102 (2.33Å), ARG104 (1.95Å), HIS139 (2.57Å)	GLY102 (3.44Å), VAL112 (4.27Å, 5.13Å), LEU117 (4.26Å), ILE134 (4.42Å)
C17	GLY102 (1.93Å), ARG104 (2.58Å), GLN116 (2.11Å), HIS139 (2.10Å)	PRO105 (5.43Å), VAL113 (4.27Å), LEU117 (4.27Å, 5.17Å), LEU131 (4.35Å), ILE134 (5.16Å)

4. Discussions

A compound's ADMET characteristics relate to how it is absorbed, distributed, metabolized, excreted, and processed by and through the human body. To assess a drug's pharmacodynamic activity, it is crucial to consider ADMET, which makes up the pharmacokinetic profile of the drug molecule (11). The computational technique named molecular docking is used to find a ligand that will fit energetically and physically at the binding site of a protein. A stronger bond between a substance and a protein is indicated by a binding affinity value that is more negative (12). Low binding affinity values also suggest that protein-ligand binding requires less energy. The first pose is always regarded as the ideal pose since it has the highest binding affinity and the last pose has the lowest binding affinity to the target protein (13).

All the 19 derivatives were studied for ADMET analysis and found that only 8 compounds were shown to retain the Lipinski rule and no toxicity has been reported. Further, the screened 8 compounds were subjected to docking study and visualization of ligand interactions. In comparison to the binding affinity of the co-crystal inhibitor, 6 tetracycline derivatives have exhibited a greater affinity for chain A of the Tet R protein. These 6 variants include C1 with fluoro substituent at R4 position (-8.9 kcal/mol), C11 with methyl group at R4 (-8.4kcal/mol), C12 with ethyl group at R4 (-8.5 kcal/mol), C14 with isopropyl substituent at R4 (-7.7 kcal/mol), C16 with ethenyl substituent at R4 (-7.7 kcal/mol), C17 with ethynyl substituent at R4 (-8.6 kcal/mol). These substances are further studied for the visualization of ligand interactions. It can be seen from the ligand interactions in figure 4 and table 5 that all 6 compounds have formed traditional H-bonds with various amino acids in the active site of the Tet R protein. Among these 6 tetracycline derivatives, C1 is found to have a better binding affinity towards the target protein.

Researchers now apply to trim methods that differ from the traditional approaches used to examine synthetic medicines. In-silico techniques like molecular docking and molecular dynamics simulations have been employed more and more in drug discovery research to locate promising synthetic compounds for the treatment of various ailments. But certain medications have poor oral bioavailability. To address the issues with drug bioavailability, many researchers have adopted novel drug delivery strategies. Scientific developments have led to the use of novel techniques in drug discovery programs by pharmaceutical researchers (14).

5. Conclusions

The results of the current in-silico analysis indicate that six compounds can inhibit the Tet R protein of the tetracycline resistance element. Compared to the co-crystal inhibitor, they have shown a greater binding affinity for the Tet R's active binding site. Among the six compounds, Compound 1 (Fluorine at 7th position) has shown a better binding affinity towards the target protein. Additionally, these molecules have a favorable ADME profile and no toxicity. To effectively evaluate the compound's inhibitory potential against Tet R protein of the tetracycline resistance element, however, additional molecular dynamic simulation study and (in-

vitro/in-vivo) research is required as the current work is limited to only molecular docking and ADMET in-silico model.

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Conflict of Interest

The authors declare no conflicting interest.

Authors contribution

All the authors have made equal contributions in carrying out the work.

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