



# Isolation, Characterization, and Identification of Yeast from Fermented Catfish (*pangasius Sp.*) Sausage

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**Abstract:** Fermented sausage is a product that results from the work of lactic acid-forming bacteria, both those naturally present in meat and added starter bacteria. In addition to lactic acid bacteria, other microorganisms such as yeast are also found in the fermentation process. The purpose of this study was to identify the yeast genus isolated from fermented patin fish sausage. The methods used in this study were exploratory, namely descriptive identification of yeast (morphological and biochemical tests) and quantitative tests for pH, Aw, total yeast, and organoleptic properties in sausages fermented spontaneously for 6 days. Based on morphological and biochemical testing for yeast identification, it was found that the type of yeast found in fermented patin fish sausage was similar to the genus *Candida* sp. Fermented patin fish sausage was well fermented, as seen from the organoleptic quality values produced, including a brownish-red appearance with a strong smoky aroma, a delicious and spicy taste, the characteristic acidity of fermented products, and no fishy smell, as well as a compact, dense, and elastic texture. This fermented patin fish sausage also has a pH value of 4.23, Aw: 0.62, and Total yeast:  $2.6 \times 10^4$  CFU/mL. Lactic acid bacteria (LAB) are not the only microorganisms involved in the fermentation process, as yeast is also found and has been proven to contribute to the quality of fermented catfish sausage.

## Introduction

Fermented sausage is a product that results from the work of lactic acid-forming bacteria, either naturally present in meat or added as starter bacteria. Fermented sausage is made from a mixture of raw meat, fat, and other ingredients, which is then stuffed into casings and left to ferment and mature (1). The fermentation process involving microbial activity is carried out for various purposes, including to increase product shelf life, create distinctive flavor and aroma characteristics, and enhance nutritional value to produce functional food products (2). However, in addition to lactic acid bacteria (LAB), other microorganisms such as yeast are also found in the fermentation process and are thought to contribute to product quality. Yeast is one of the microorganisms used as an inoculum or starter in the fermentation process to produce certain products (3). Yeast in the fermentation process of patin fish sausage produces compounds that contribute to the aroma, flavor, and texture of the sausage. The acidic conditions formed during the sausage fermentation process affect the sensory characteristics, particularly the taste and texture of the product. The distinctive sour taste (tangy) is produced as a result of

fermentation activity by yeast, giving fermented sausages their unique taste (4).

The dominant yeasts commonly found in fermented foods (fish and meat) are from the genera *Candida*, *Saccharomyces*, *Pichia*, *Zygosaccharomyces*, *Debaryomyces*, and *Hanseniaspora* (5). Yeast capable of breaking down proteins (having proteolytic activity) is thought to play an important role in the fermentation of fish such as belacan depik and fish paste (6). According to (7), yeast has various beneficial effects on human health and shows potential as a probiotic microorganism. The presence of yeast in fermented sausages affects product quality characteristics, including color, taste, and aroma, and has the ability to degrade peroxide compounds and exhibit lipolytic and proteolytic activity.

Several studies have attempted to explain the role of yeast as secondary microbiota in fermented products. Identification using classical methods of yeast isolated from sheep meat products showed the presence of several genera such as *Debaryomyces*, *Candida*, *Yarrowia*, *Pichia*, *Rhodotorula*, *Cryptococcus*, and *Trichosporon* (8). Evaluation of yeast diversity during sausage processing and maturation in various factories showed the presence of *Candida* and *Trichosporon* species in raw meat or in the

early stages of maturation, while *Candida zeylanoides*, *Yarrowia lipolytica*, and *Debaryomyces hansenii* appeared in the final stages of sausage production (9). In the sausage fermentation process, lactic acid bacteria (LAB), which are commonly used as starters, have been extensively studied. However, the role of other microorganisms, such as yeast in sausage fermentation, is still not fully understood. Based on the above description, the author was interested in conducting research on the isolation, characterization, and identification of yeast in fermented patin fish sausage.

## Methodology

### Time and Places

This research was conducted from May 2025 to June 2025 at the Fisheries Product Processing Laboratory and the Fisheries Product Microbiology Laboratory, Faculty of Fisheries and Marine Sciences, University of Riau.

### Materials

The ingredients used in making fermented sausage are catfish (*Pangasius sp.*) obtained from traditional markets, polyamide casings with a diameter of 1.9, tapioca flour, carrageenan, isolate soy protein (ISP), salt, pepper (*Piper nigrum L.*), garlic (*Allium sativum L.*), red yeast rice (*Monascus purpureus*), skim milk, corn oil, frankfurters, bratwurst, smoke powder, and ice cubes. The materials used for yeast isolation, characterization, and identification testing were PDA (Potato Dextrose Agar), chloramphenicol (100 µg/mL), distilled water, 70% alcohol, physiological solution (0.9% NaCl), crystal violet, iodine, 95% ethanol, 0.5% safranin, malachite green 0.5%, CuSO solution 1%, immersion oil, Christensen's Urea Broth (1g peptone, 5g NaCl, 2g H<sub>2</sub>P<sub>4</sub>, 12µg red phenol), and 20% sterile urea).

The equipment needed to make fermented sausage includes containers, trays, coconut shells, sausage molds, mattress thread, stoves, boiling pots, scissors, and knives. Microbiological analysis uses equipment such as masks, gloves, Bunsen burners, mortars, cotton wool, label paper, aluminum foil, Erlenmeyer flasks, test tubes, Petri dishes, micropipettes, tips, measuring flasks, incubators, hot plates, autoclaves, object glasses, cover glasses, test tubes, ose nedles, Bunsen burners, and microscopes.

## Research method

This Study used an exploratory approach, which aims to isolate and identify the types of yeast (genus level) found in fermented patin fish sausage. The yeast identification results were analyzed and presented in tables and figures to clearly visualize the data.

Sampling was conducted on day 0 (before fermentation) and day 6 (final fermentation stage). At each sampling time, pH, water activity (A<sub>w</sub>), and total yeast counts were measured.

## Research procedure

### Making fermented catfish sausage

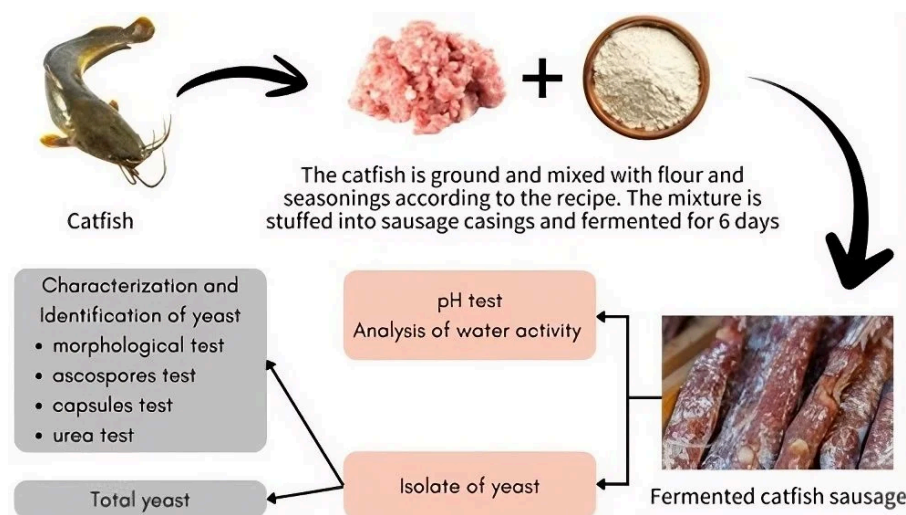
In the production of sausages, fresh catfish undergo a series of cleaning processes, including gutting, removal of the intestines and gills, and washing with running water. After that, the fish is cut into fillets. The catfish is minced for 60 seconds, then the prepared seasoning is added. The grinding process continues until the mixture reaches the desired consistency (smooth). Next, the mixture is stuffed into 1.9 cm diameter polyamide casings with a sausage length of 7-10 cm. The fish sausages are steamed at 40°C for 30 minutes, followed by a 24- hour conditioning process at room temperature. The next process involves baking at a temperature between 40°C and 45°C for 2 days, with each smoking session lasting 3 hours. The final stage is fermenting the sausage for 6 days (10).

### Yeast Isolation

Samples of fermented catfish sausage were cultured using Total Plate Count on a medium (Potato Dextrose Agar) containing 100 µg/mL chloramphenicol (to inhibit bacterial growth), then incubated for 48 hours at 37°C. Colonies suspected to be yeast were observed morphologically under a microscope and subcultured on PDA medium and stored at 4°C (11). The overall experimental workflow, including sausage fermentation, physicochemical analysis, yeast isolation, and identification, is presented in **Figure 1**.

### Water activity (A<sub>w</sub>)

Water activity (A<sub>w</sub>) was measured on days 0 and 6 of fermentation using a pre-calibrated A<sub>w</sub> meter. A total of 4



**Figure 1.** Research process on fermented catfish sausage.

g of crushed fermented sausage was placed in a measuring cup. The device was used until it showed the "completed" sign (12).

### pH test

pH measurement was conducted on days 0 and 6 of fermentation using a calibrated pH meter. A total of 10 g of sausage sample was homogenized with 90 mL of distilled water to obtain a uniform suspension. The pH meter was calibrated using standard buffer solutions at pH 4.0 and 7.0 prior to measurement. The electrode was immersed in the homogenized sample, and the pH value was recorded once a stable reading was obtained (13).

### Total yeast

Total yeast counts were determined on days 0 and 6 of fermentation. Total yeast testing begins with dilution, by homogenizing 10 g of fermented sausage sample. This homogenate is a  $10^{-1}$  dilution. To obtain a  $10^{-2}$  dilution, 1 mL of the  $10^{-1}$  dilution is taken and added to a sterile reaction tube containing 9 mL of physiological saline, then homogenized. Continue the dilution process up to a  $10^{-5}$  dilution. Then, transfer 1 mL of sample from the last four dilution series ( $10^{-3}$ ,  $10^{-4}$ , and  $10^{-5}$ ) into Petri dishes and add 15 mL of PDA medium. Next, the petri dishes are incubated at 37°C for 3 days. After that, count the yeast that have grown on the petri dishes using the Eq. 1:

### Identification and Characterization of Yeast at the Genus Level

#### Macroscopic and microscopic morphological observations

Macroscopic observation of yeast characteristics refers to the guidebook *The Yeast Taxonomic Study* (15) The morphological characteristics observed are based on the growth of colonies visible on solid media, which include the surface of the colony, the edge or margin of the colony, the texture of the colony, the color of the colony, and the elevation (16). Microscopic observation of yeast morphology was carried out by making a wet preparation,

namely by placing the purified yeast isolate 3x24 hours on an object glass, then dripping sterile distilled water (17). The next step is observation under a microscope with a magnification of 4x10 to 100x10 to observe the size of the yeast, budding, and cell shape (18). The observation results refer to the book *The Yeast: A Taxonomy Study* by (15).

### Ascospore test

The ascospore test was performed using a modified Schaeffer-Fulton technique with 0.5% malachite green and 0.5% safranin in a 1: 1 ratio. One drop of the isolate was stained with these dyes, then observed under a microscope. Mature ascospores appeared green, while vegetative yeast cells appeared red (19).

### Capsule test

Capsule testing on yeast is performed using the smear method on a test slide. One ose of the isolate is placed on the test slide, which is then stained with crystal violet for  $\pm$  5 minutes and rinsed with a 1% CuSO<sub>4</sub> solution. Observation is performed under a microscope at 1000x magnification with the addition of immersion oil. The capsule will appear as a faint blue circle surrounding the purple-colored vegetative yeast cells (19).

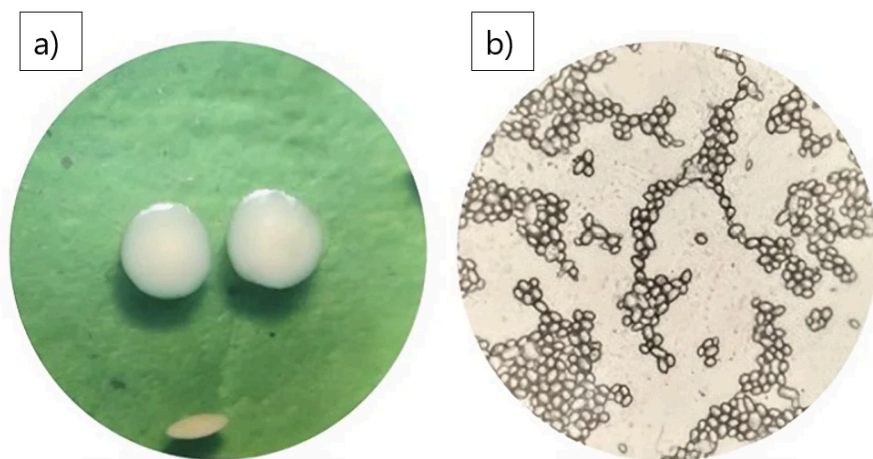
### Urea test

The isolated yeast strain was inoculated into Christensen's Urea Broth medium, which consisted of a mixture of 1 g peptone, 5 g NaCl, 2 g H<sub>2</sub>PO<sub>4</sub>, and 12  $\mu$ g phenol red dissolved in 1 L of sterile distilled water with a pH of 6.8. A total of 4.5 mL of the medium solution was transferred to a test tube and sterilized using an autoclave for 15 minutes. A total of 0.5 mL of sterile 20% urea was added to each tube. The strain was incubated at 25°C for 4 days and checked daily. A positive result of this test is the formation of a pink color on the medium in the tube (20).

### Data Analysis

The data obtained is presented in tables and graphs to facilitate understanding and interpretation. Furthermore, the data is analyzed descriptively with reference to

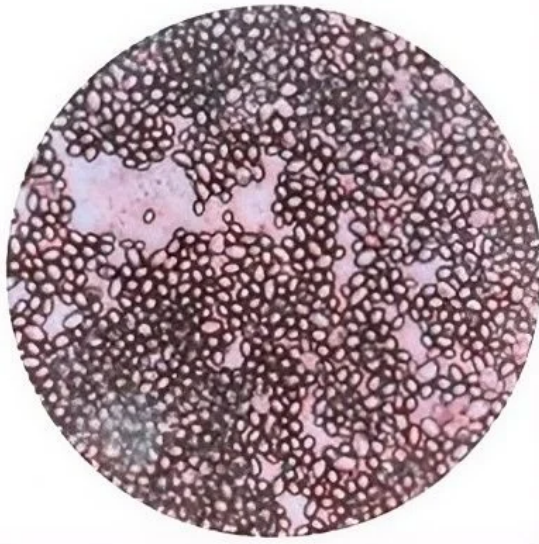
$$N = \frac{\sum C}{[(1 \times n_1) + (0.1 \times n_2)] \times d} \quad (\text{Eq. 1})$$



**Figure 2.** Morphological observation; a) Macroscopic, b) Microscopic



**Figure 3.** Ascospore test observation.



**Figure 4.** Capsule test observation.



**Figure 5.** Urease test observation

relevant literature studies to gain deeper insight into the findings.

## Result and Discussion

### Morphology of Yeast

Morphological observations were conducted macroscopically and microscopically on the isolates obtained. Morphological observations of the colonies included colony shape, edges, elevation, and color. Meanwhile, morphological observations of the cells were conducted to determine the cell shape of the yeast isolates. The results of the yeast morphological observations are presented in **Figure 2**.

Based on **Figure 2**, macroscopic observations show that yeast colonies are milky white, evenly elevated, shiny with smooth and even edges. The same is reported by (21), that yeast isolates generally have a milky white color, and some yeast isolates have a creamy white, yellowish white, and brownish white color. Yeast isolates have a flat and raised elevation, all isolates have a shiny surface with smooth and flat edges. Microscopic observations show that yeast cells are round, oval, and without pseudohyphae. Based on the microscopic similarity of the isolate, it is suspected to belong to the *Candida* genus of yeast. This is supported by the opinion of (22), that yeast of the *Candida* genus have cell shapes that vary from round, oval, cylindrical to elongated, rarely apiculate, ogival, triangular or bottle-shaped with or without pseudohyphae. Asexual reproduction occurs through multilateral budding. However, to confirm that the isolated genus is indeed *Candida*, biochemical testing is required to ensure the yeast identification process is more accurate and clear.

### Biochemical Characteristics of Yeast

#### Ascospore Test

Based on **Figure 3**, the results of the ascospore test for the genus *Candida* were negative. No ascoporation structures were visible after staining, because *Candida* is an anascoprogenic yeast that is unable to form asci or ascospores naturally (20). This is in line with the opinion of (22) that the genus *Candida* does not form ascospores, arthrospores, teliospores, or blastospores, but chlamydospores may form in some species that do not have carotenoid pigments and are therefore white to cream in color.

#### Capsule Test

Based on **Figure 4**, the capsule staining test results were negative. No halo or capsule structure was detected, because *Candida* does not produce polysaccharide capsules (20). Capsule staining tests on yeast of the *Candida* genus have been conducted by (22), where based on microscopic observations, no capsules in the form of pale blue circles surrounding purple yeast cells were observed in the isolates examined.

#### Urease Test

Based on **Figure 5**, the urease test results for the *Candida* genus were negative. There was no change in the color of the medium to pink after incubation, indicating no urea hydrolysis (17). The absence of urease activity in this isolate supports that the genus found is consistent with the characteristics of *Candida*, which relies more on other metabolic mechanisms to survive and thrive in its host

environment (23). This negative result distinguishes it from pathogenic yeasts such as *Candida albicans*, which are urease-positive and contribute to virulence in lung tissue (24). Urease testing on yeasts of the genus *Candida* has been conducted by (16), in which the yeast isolates tested showed no reaction of media color change to pink.

### Chemical and Microbiological Analysis

Based on **Table 1**, it is known that fermented sausage undergoes changes in pH value during the fermentation process. Several factors that influence the decrease in pH are due to the smoking process. According to (25), pH can decrease during the smoking process because the smoke components that adhere to the sausage have an acidic taste, such as carboxylic acids, which include formic acid, acetic acid, and butyric acid. On day 0, fermented sausage was found to have a pH value of around 6. On the sixth day, the pH value of the sausage decreased to around 4. This was due to lactic acid bacteria beginning to work and producing acids that lowered the pH. According to (26), the longer the fermentation, the greater the decrease in pH due to an increase in the number of lactic acid bacteria, which also increases acid production. According to (27), this decrease in pH can also be caused by the growth of lactic acid bacteria, resulting in a lower pH of the substrate.

In addition, other factors include additives such as skim milk and liquid smoke. Skim milk can lower pH because it contains lactose, which can stimulate the growth of lactic acid bacteria, leading to fermentation (28). Liquid smoke can lower the pH value of a product (29). The pH value of liquid smoke ranges from 1.50 to 4.50, with quality classification 1 having a pH of 1.50–2.75 and quality 2 having a pH of 2.76–4.50. This value indicates the acidic nature of liquid smoke, which comes from the content of organic acid compounds such as acetic acid and formic acid (30).

### Water Activity (Aw)

Based on **Table 1**, it is known that fermented sausage undergoes changes in water activity. The decrease in Aw value is also thought to be caused by the evaporation process due to smoking carried out before the fermentation process (10). The smoking process causes the surface of the sausage to become drier due to the evaporation of free water. The decrease in water activity (Aw) is caused by the drying process, which inhibits the growth of spoilage microorganisms (31). However, the Aw level must be maintained so that the microorganisms that act as fermenters can work properly. Microbes can generally grow on food materials with Aw 0.6–0.99 (32). This decrease in Aw is thought to be related to a decrease in product pH due to acids produced by yeast activity,

causing proteins to lose their ability to retain water, resulting in dry fermented sausages (33). The longer the fermentation process lasts, the drier the fermented sausage will be and the better its appearance. This is supported by the opinion of (34) that a decrease in Aw helps reduce microbial growth and chemical reactions that damage materials, such as hydrolysis, browning, and fat oxidation.

### Total Yeast

Based on the data in **Table 1**, it can be seen that the amount of yeast will increase as the fermentation time increases. This is in line with the research by (35), which found that fermentation time greatly affects yeast activity because the longer the fermentation, the greater the amount of yeast or the more active the yeast is in reproducing. Yeast can grow optimally under suitable conditions. Yeast growth factors can be influenced by the availability of nutrients, both macronutrients and micronutrients. The nutritional composition of the medium is very important to support yeast growth and metabolism (36). Nutrients in the growth medium will be used for growth, resulting in an increase in the number of yeast cells. 256 Another factor that also influences this is the length of time the isolate culture is stored, which can cause the nutrients in the medium to be completely consumed, leading to the death of the isolate in the storage medium. This can cause the isolate to be unable to grow in a new culture. In addition, it can also be caused by insufficient nutrients in the growth medium (37).

### Conclusion

The conclusions of this study indicate that the yeast isolated from fermented catfish sausage belongs to the genus *Candida* sp., which exhibits diverse morphological characteristics, including round, oval, cylindrical, and elongated shapes, and may present with or without pseudohyphae. Biochemical characterization showed that *Candida* sp. does not form ascospores, does not possess capsules, and does not produce urea. The presence and activity of yeast were found to influence the chemical and microbiological properties of the fermented product, as reflected in an average pH value of 4.23, water activity (Aw) of 0.62, and total yeast count of  $2.6 \times 10^4$  CFU/mL. Furthermore, the results demonstrate that lactic acid bacteria (LAB) are not the only microorganisms involved in the fermentation process, as yeast also plays a significant role in contributing to the overall quality of the product. Consequently, these findings highlight the complex microbial synergy required to develop the unique flavor profiles and structural integrity characteristic of traditional fermented sausages.

**Table 1.** Average pH, Aw, and total yeast values of fermented sausage

| Parameter   | Average  |                             |
|-------------|----------|-----------------------------|
|             | Day - 0  | Day - 6                     |
| pH          | 6, 70    | 4, 23                       |
| Aw          | 0, 67    | 0, 62                       |
| Total yeast | 0 CFU/mL | 2, 6x10 <sup>4</sup> CFU/mL |

## Declaration

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### Conflict of Interest

The authors declare no conflicting interest.

### Data Availability

The unpublished data will be provided upon request to corresponding author.

### Ethics Statement

Not applicable.

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## Additional Information

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